

## Two New Sesquiterpenes from *Euonymus alatus*

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Two new sesquiterpenes, **1** and **2**, as well as the five known compounds **3**–**7**, were isolated from the stems of *Euonymus alatus*. Compounds **2**–**7** have a  $\beta$ -dihydroagarofuran skeleton. The structures of these compounds were elucidated mainly by spectroscopic methods (1D-, 2D-NMR, ESI-MS, and HR-ESI-MS). We also report the X-ray crystal structure of evonine (**3**) for the first time.

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**Introduction.** – The Celastraceae, which contain about 98 genus and 1210 species [1], produce various  $\beta$ -dihydroagarofuran sesquiterpene polyol esters and alkaloids [2]. All these constituents belong to the eudesmane sesquiterpene family. Meanwhile, these kinds of compounds have also been isolated from the plants of the genus *Euonymus* [3–10]. *Euonymus alatus*, known as winged euonymus or ‘Gui Jian Yu’ in China in the genus *Euonymus*, has been used as a folk medicine for over 2000 years [11] and is widely used for the therapy of tumors, diabetes, and wound [12–16]. So far, several kinds of compounds have been isolated from this plant, including cardenolides, flavonoids, steroids, triterpenes, and sesquiterpenes [14][16–21]. Our phytochemical studies led to the isolation of the two new sesquiterpenes **1** and **2**, along with five known ones, **3**–**7**, from the 95% EtOH extract of the stems of *Euonymus alatus*. We also report here the X-ray crystal structure of evonine (**3**) for the first time.

**Results and Discussion.** – The 95% EtOH extract of the stems of *E. alatus* was partitioned into fractions soluble in petroleum ether and AcOEt. Repeated column chromatography over silica gel (SiO<sub>2</sub>), *Sephadex LH-20*, and *ODS* of the petroleum ether fraction led to seven compounds, *i.e.*, the two new sesquiterpenes **1** and **2** (Fig. 1), along with five known ones evonine (**3**), neoevonine (**4**), 1 $\beta$ ,2 $\beta$ ,5 $\alpha$ ,8 $\beta$ ,11-pentaacetoxy-4 $\alpha$ -hydroxy-3 $\alpha$ -(2-methylbutanoyl)-15-nicotinoyl-7-oxo-dihydroagarofuran (**5**), 6 $\alpha$ ,12-diacetoxy-2 $\beta$ ,9 $\alpha$ -di( $\beta$ -furancarboxyloxy)-4 $\alpha$ -hydroxy-1 $\beta$ -(2-methylbutanoyloxy)- $\beta$ -dihydroagarofuran (**6**), 1 $\alpha$ ,2 $\alpha$ ,6 $\beta$ -triacetoxy-4 $\beta$ -hydroxy-9 $\beta$ -( $\beta$ -furancarboxy-15-[( $\alpha$ -methyl)butyroyloxy]- $\beta$ -dihydroagarofuran (**7**).

Compound **1**<sup>1</sup>), a colorless oil, showed the  $[M + Na]^+$  ion at  $m/z$  361.2345 in its HR-ESI-MS, suggesting the molecular formula C<sub>20</sub>H<sub>34</sub>O<sub>4</sub> and the presence of 4 degrees of

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<sup>1</sup>) Trivial atom numbering; for systematic names of **1** and **2**, see *Exper. Part*.

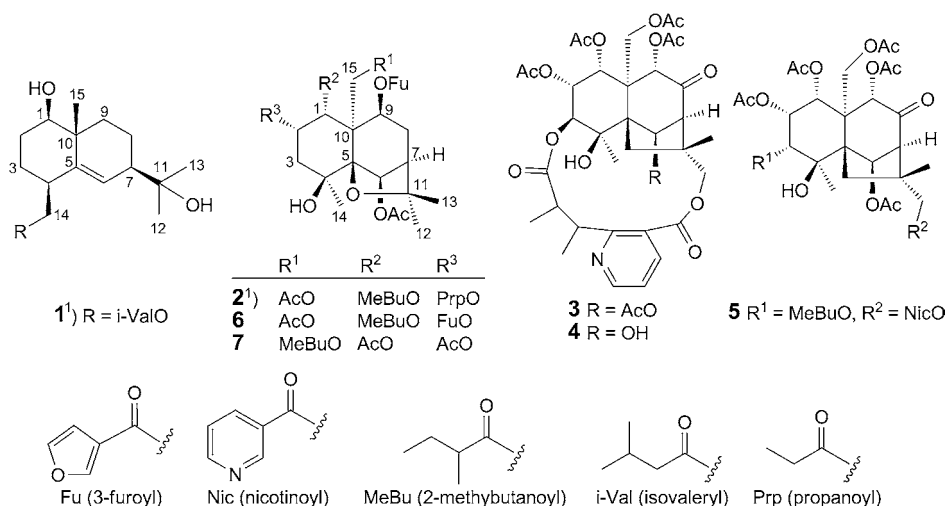


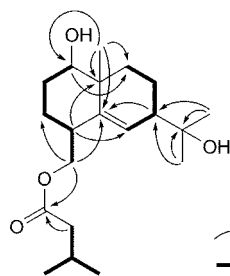
Fig. 1. Compounds **1**–**7**, isolated from *Euonymus alatus*

unsaturation. The IR spectrum showed OH-type absorption bands at  $3436\text{ cm}^{-1}$  and ester-carbonyl bands at  $1733\text{ cm}^{-1}$ . The  $^1\text{H-NMR}$  spectrum (Table 1) revealed two *ds* at  $\delta(\text{H})$  0.92 ( $J = 6.8\text{ Hz}$ , 6 H) and 2.15 ( $J = 6.4\text{ Hz}$ , 2 H) and one *m* at  $\delta(\text{H})$  2.01–2.06 (overlapped, 1 H), consistent with the presence of an isovaleryloxy (= 3-methyl-1-oxobutoxy) unit. Moreover, the  $^1\text{H-NMR}$  spectrum showed three *Me s* at  $\delta(\text{H})$  0.96, 1.11, and 1.13, one  $\text{CH}_2$  group at  $\delta(\text{H})$  3.98 (*dd*,  $J = 6.8, 10.8\text{ Hz}$ , 1 H) and 4.23 (*dd*,  $J = 9.2, 10.8\text{ Hz}$ , 1 H), and the H-atom of a trisubstituted double bond at  $\delta(\text{H})$  5.66 (*d*,  $J = 2.8\text{ Hz}$ , 1 H), the latter being also suggested by the  $^{13}\text{C-NMR}$  signals (Table 1) at  $\delta(\text{C})$  144.37 (*s*) and 129.24 (*d*). The HSQC and COSY data (Table 1 and Fig. 2) established the fragments  $\text{CH}(1)\text{CH}_2(2)\text{CH}_2(3)\text{CH}(4)\text{CH}_2(14)$  and  $\text{CH}(6)\text{CH}(7)\text{CH}_2(8)\text{CH}_2(9)$ . The HMBC experiment (Fig. 2) and all the above data presented the skeleton fragment correlation from C(1) to C(10). In the HMBC spectrum, the cross-peaks  $\delta(\text{H})$  0.96 (*s*, Me(15))/ $\delta(\text{C})$  78.22 (C(1)), 35.99 (C(9)), and 41.12 (C(10)),  $\delta(\text{H})$  1.11 (*s*, Me(12)) and 1.13 (*s*, Me(13))/ $\delta(\text{C})$  74.03 (C(11)) and 47.36 (C(7)), as well as  $\delta(\text{H})$  3.98 and 4.23 (*2dd*,  $\text{CH}_2(14)$ )/ $\delta(\text{C})$  174.95 (C=O of isovaleryloxy) were present. Moreover, the molecular mass and the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data indicated that C(1) ( $\delta(\text{C})$  78.22) and C(11) ( $\delta(\text{C})$  74.03) should be substituted by OH groups. In the NOESY experiment (Fig. 3), H–C(1) ( $\delta(\text{H})$  3.25–3.29) showed a correlation with H–C(4) ( $\delta(\text{H})$  2.49–2.54); thus both H-atoms were arbitrarily assigned as being on the  $\alpha$ -side. The NOE correlation Me(15) ( $\delta(\text{H})$  0.96)/ $\text{CH}_2(14)$  ( $\delta(\text{H})$  3.98 and 4.23) and the absence of a correlation Me(15)/H–C(1) indicated that Me(15) and C(14) were on the  $\beta$ -side. The NOEs  $\text{H}_\alpha\text{-C}(9)$  ( $\delta(\text{H})$  1.65–1.67)/H–C(1) and H–C(7) ( $\delta(\text{H})$  2.01–2.06) showed that H–C(7) was on the  $\alpha$ -side. Thus the relative configuration of **1** was determined. All the above data and comparison with those given in [22] established the structure of **1**.

Compound **2**<sup>1)</sup> was obtained as a white powder. Its molecular formula  $\text{C}_{32}\text{H}_{44}\text{O}_{13}$  was deduced from the HR-ESI-MS ( $m/z$  659.2706 ( $[M + \text{Na}]^+$ )), indicating the presence of 11 degrees of unsaturation. The IR spectrum showed an OH-type absorption band at

Table 1.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data (400 and 100 MHz, resp.;  $\text{CD}_3\text{OD}$ ) of **1**).  $\delta$  in ppm,  $J$  in Hz.

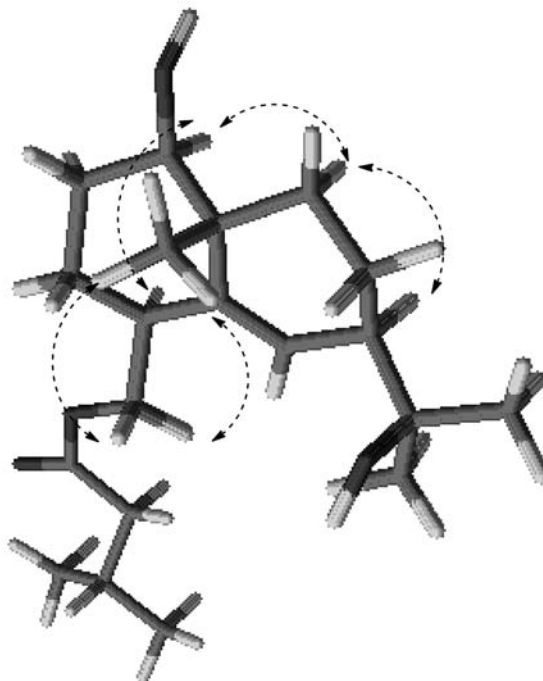
Position	$\delta(\text{H})$	$\delta(\text{C})$	$^1\text{H}, ^1\text{H}$ -COSY
H–C(1)	3.25–3.29 ( <i>m</i> , overlapped)	78.2 ( <i>d</i> )	H–C(2)
CH <sub>2</sub> (2)	1.56–1.61 ( <i>m</i> )	27.5 ( <i>t</i> )	H–C(1), H <sub><math>\alpha</math></sub> and b–C(3)
CH <sub>2</sub> (3)	1.70–1.75 ( <i>m</i> , H <sub><math>\alpha</math></sub> ), 1.48–1.51 ( <i>m</i> , H <sub><math>\beta</math></sub> )	27.1 ( <i>t</i> )	H–C(2), H <sub><math>\beta</math></sub> –C(3), H–C(4) H–C(2), H <sub><math>\alpha</math></sub> –C(3), H–C(4)
H–C(4)	2.49–2.54 ( <i>m</i> )	45.1 ( <i>d</i> )	H <sub><math>\alpha</math></sub> and b–C(3), H <sub><math>\alpha</math></sub> and b–C(14)
C(5)		144.3 ( <i>s</i> )	
H–C(6)	5.66 ( <i>d</i> , $J=2.8$ )	129.2 ( <i>d</i> )	H–C(7)
H–C(7)	2.01–2.06 ( <i>m</i> , overlapped)	47.3 ( <i>d</i> )	H–C(6), H <sub><math>\alpha</math></sub> and $\beta$ –C(8)
CH <sub>2</sub> (8)	1.54–1.56 ( <i>m</i> , H <sub><math>\alpha</math></sub> ), 1.62–1.64 ( <i>m</i> , H <sub><math>\beta</math></sub> )	21.2 ( <i>t</i> )	H–C(7), H <sub><math>\beta</math></sub> –C(8), H <sub><math>\alpha</math></sub> and $\beta$ –C(9) H–C(7), H <sub><math>\alpha</math></sub> –C(8), H <sub><math>\alpha</math></sub> and $\beta$ –C(9)
CH <sub>2</sub> (9)	1.65–1.67 ( <i>m</i> , H <sub><math>\alpha</math></sub> ), 1.45–1.47 ( <i>m</i> , H <sub><math>\beta</math></sub> )	35.9 ( <i>t</i> )	H <sub><math>\alpha</math></sub> and $\beta$ –C(8), H <sub><math>\beta</math></sub> –C(9) H <sub><math>\alpha</math></sub> and $\beta$ –C(8), H <sub><math>\alpha</math></sub> –C(9)
C(10)		41.1 ( <i>s</i> )	
C(11)		74.0 ( <i>s</i> )	
Me(12)	1.11 ( <i>s</i> )	27.2 ( <i>q</i> )	
Me(13)	1.13 ( <i>s</i> )	27.5 ( <i>q</i> )	
CH <sub>2</sub> (14)	4.23 ( <i>dd</i> , $J=9.2, 10.8$ , H <sub><math>\alpha</math></sub> ), 3.98 ( <i>dd</i> , $J=6.8, 10.8$ , H <sub><math>\beta</math></sub> )	67.8 ( <i>t</i> )	H–C(4), H <sub><math>\beta</math></sub> –C(14) H–C(4), H <sub><math>\alpha</math></sub> –C(14)
Me(15)	0.96 ( <i>s</i> )	21.0 ( <i>q</i> )	
i-ValO		174.9 ( <i>s</i> )	
	2.15 ( <i>d</i> , $J=6.4$ )	44.5 ( <i>t</i> )	
	2.01–2.06 ( <i>m</i> , overlapped)	27.0 ( <i>d</i> )	
	0.92 ( <i>d</i> , $J=6.8$ )	22.9 ( <i>q</i> )	
	0.92 ( <i>d</i> , $J=6.8$ )	22.9 ( <i>q</i> )	



HMBC  
 $^1\text{H}, ^1\text{H}$ -COSY

Fig. 2. Selected 2D-NMR correlations of **1**

3445  $\text{cm}^{-1}$  and ester-carbonyl bands at 1749 and 1718  $\text{cm}^{-1}$ . The  $^1\text{H}$ -NMR spectrum (Table 2) exhibited a *s* ( $\delta(\text{H})$  7.97 (1 H)) and two *d* ( $\delta(\text{H})$  6.68 ( $d$ ,  $J=1.0$  Hz, 1 H) and 7.37 ( $d$ ,  $J=1.0$  Hz, 1 H)), which were attributed to a (furylcarbonyl)oxy substituent. A *t* ( $\delta(\text{H})$  0.63 ( $J=7.5$  Hz, 3 H)), a *d* ( $\delta(\text{H})$  0.84 ( $J=7.5$  Hz, 3 H)), and three *m* ( $\delta(\text{H})$  1.91–1.97, 1.24–1.32, and 0.98–1.07 (1 H each)) were attributed to a (2-methylbutanoyl)oxy substituent, and a *t* ( $\delta(\text{H})$  1.12 ( $J=7.5$  Hz, 3 H)) and a *q* ( $\delta(\text{H})$  2.33 ( $J=7.5$  Hz, 2 H)) to a propanoyloxy moiety. All these data were certified by the  $^{13}\text{C}$ -NMR spectrum (Table 2). Moreover, the  $^1\text{H}$ -NMR spectrum contained signals assignable to four CH groups carrying an ester group at  $\delta(\text{H})$  6.07 (*s*, H–C(6)), 5.54 (*d*,  $J=3.0$  Hz, H–C(1)), 5.45–5.47 (*m*, H–C(2)), and 5.17 (*d*,  $J=7.0$  Hz, H–C(9)), to a

Fig. 3. Key NOESY correlations of **1**

CH<sub>2</sub> group carrying an ester group at  $\delta(\text{H})$  4.35 and 4.95 (*AB*,  $J = 13.0$  Hz, CH<sub>2</sub>(15)), to a tertiary Me group at  $\delta(\text{H})$  1.41 (*s*, Me(14)) attached to a C-Atom at  $\delta(\text{C})$  69.80 (C(4)) carrying an OH group, to two acetate Me groups at  $\delta(\text{H})$  2.07 and 2.22 (*2s*), and to two tertiary Me groups at  $\delta(\text{H})$  1.43 (*s*, Me(12)) and 1.50 (*s*, Me(13)). All the above data and 2D-NMR experiments (HSQC and COSY (Fig. 4)) indicated that **2** was a  $\beta$ -dihydroagarofuran sesquiterpene with one (furylcarbonyl)oxy, one (2-methylbutanoyl)oxy, one propanoyloxy, and two acetoxy groups. The positions of these substituent groups were deduced from the HMBC data (Fig. 4), which showed the correlations H–C(1) ( $\delta(\text{H})$  5.54/ $\delta(\text{C})$  174.54 (C=O of MeBuO), H–C(2) ( $\delta(\text{H})$  5.45–5.47)/ $\delta(\text{C})$

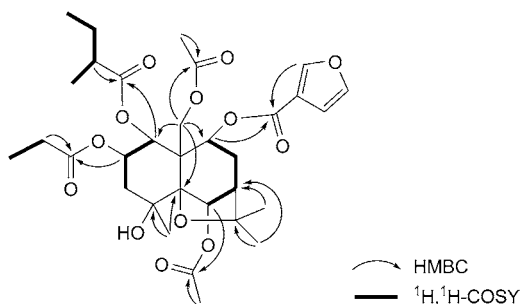
Fig. 4. Selected 2D-NMR correlations of **2**

Table 2.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data (500 and 125 MHz, resp.;  $\text{CD}_3\text{OD}$ ) of **2**<sup>1</sup>.  $\delta$  in ppm,  $J$  in Hz.

Position	$\delta(\text{H})$	$\delta(\text{C})$
H–C(1)	5.54 ( <i>d</i> , $J=3.0$ )	69.8 ( <i>d</i> )
H–C(2)	5.45–5.47 ( <i>m</i> )	68.0 ( <i>d</i> )
$\text{CH}_2(3)$	2.10 ( <i>dd</i> , $J=3.5, 15.5, \text{H}_a$ ), 1.91–1.97 ( <i>m</i> , overlapped, $\text{H}_b$ )	42.0 ( <i>t</i> )
C(4)		69.8 ( <i>s</i> )
C(5)		91.2 ( <i>s</i> )
H–C(6)	6.07 ( <i>s</i> )	78.2 ( <i>d</i> )
H–C(7)	2.12–2.14 ( <i>m</i> )	49.2 ( <i>d</i> )
$\text{CH}_2(8)$	2.49 ( <i>ddd</i> , $J=3.5, 7.0, 16.0, \text{H}_a$ ), 2.21 ( <i>dd</i> , $J=2.0, 16.0, \text{H}_b$ )	34.6 ( <i>t</i> )
H–C(9)	5.17 ( <i>d</i> , $J=7.0$ )	69.2 ( <i>d</i> )
C(10)		55.0 ( <i>s</i> )
C(11)		84.6 ( <i>s</i> )
Me(12)	1.43 ( <i>s</i> )	25.7 ( <i>q</i> )
Me(13)	1.50 ( <i>s</i> )	29.5 ( <i>q</i> )
Me(14)	1.41 ( <i>s</i> )	25.1 ( <i>q</i> )
$\text{CH}_2(15)$	4.96 ( <i>d</i> , $J=13.0, \text{H}_a$ ), 4.35 ( <i>d</i> , $J=13.0, \text{H}_b$ )	65.7 ( <i>t</i> )
FuO–C(9)		161.9 ( <i>s</i> )
	7.97 ( <i>s</i> )	149.0 ( <i>d</i> )
	7.37 ( <i>d</i> , $J=1.0$ )	143.9 ( <i>d</i> )
		118.8 ( <i>s</i> )
	6.68 ( <i>d</i> , $J=1.0$ )	110.0 ( <i>d</i> )
PrpO–C(2)		173.0 ( <i>s</i> )
	2.33 ( <i>q</i> , $J=7.5$ )	28.1 ( <i>t</i> )
	1.12 ( <i>t</i> , $J=7.5$ )	9.1 ( <i>q</i> )
MeBuO–C(1)		174.5 ( <i>s</i> )
	1.91–1.97 ( <i>m</i> , overlapped)	40.9 ( <i>d</i> )
	1.24–1.32 ( <i>m</i> )	25.6 ( <i>t</i> )
	0.98–1.07 ( <i>m</i> )	
	0.84 ( <i>d</i> , $J=7.5$ )	15.8 ( <i>q</i> )
	0.63 ( <i>t</i> , $J=7.5$ )	11.4 ( <i>q</i> )
AcO–C(6)	2.07 ( <i>s</i> )	170.4 ( <i>s</i> ), 21.7 ( <i>q</i> )
AcO–C(15)	2.22 ( <i>s</i> )	170.7 ( <i>s</i> ), 21.4 ( <i>q</i> )

173.05 (C=O of PrpO), H–C(6) ( $\delta(\text{H})$  6.07)/ $\delta(\text{C})$  170.41 (C=O of AcO), H–C(9) ( $\delta(\text{H})$  5.17)/ $\delta(\text{C})$  161.95 (C=O of FuO), and  $\text{CH}_2(15)$  ( $\delta(\text{H})$  4.35 and 4.96)/ $\delta(\text{C})$  170.74 (C=O of AcO). The NOESY experiment revealed the correlations  $\delta(\text{H})$  5.54 (H–C(1))/ $\delta(\text{H})$  5.46 (H–C(2)) and 6.68 and 7.97 (FuO–C(9)),  $\delta(\text{H})$  6.07 (H–C(6))/ $\delta(\text{H})$  2.12–2.14 (H–C(7)), 1.41 (Me(14)), 1.43 (Me(12)), and 4.35 ( $\text{H}_b$ –C(15)), and  $\delta(\text{H})$  4.35 ( $\text{H}_b$ –C(15))/ $\delta(\text{H})$  5.17 (H–C(9)). Thus the relative positions of these substituent groups were determined. All these data and comparison with those given in [23] established the structure of **2**.

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### Experimental Part

*General.* Column chromatography (CC): silica gel (SiO<sub>2</sub>, 200–300 or 300–400 mesh; *Yantai Jiangyou Silica Gel Limited Company*, Yantai, P. R. China), *Sephadex LH-20* (*Pharmacia Fine Chemicals*, Piscataway, NJ, U.S.A.), and *ODS* (*Merck*, Germany). TLC and prep. TLC: *HSGF-254* silica gel plates (SiO<sub>2</sub>, 10–40 μm; *Yantai Jiangyou Silica Gel Limited Company*, Yantai, P. R. China). Optical rotations: *Perkin-Elmer-341* digital polarimeter. UV Spectra: *Shimadzu-UV-2550* spectrophotometer; λ<sub>max</sub> in nm. IR Spectra: *Bruker-Vector-22* spectrophotometer; KBr pellets; ν̄ in cm<sup>-1</sup>. NMR Spectra: *Varian-500* (500 MHz) and *Bruker-Avance-400* spectrometers; δ in ppm rel. to Me<sub>4</sub>Si as internal standard, *J* in Hz. MS: *Agilent-1100-LC/MSD-Trap* (ESI) and *Agilent-6520 Q-Tof* (HR-ESI) spectrometers; in *m/z*.

*Plant Material.* The stems of *Euonymus alatus* were collected from Kunming, Yunnan Province, P. R. China, in May 2009 and authenticated by Professor *Li-Shan Xie* of the Kunming Institute of Botany. A voucher specimen (No. 200905003) was deposited with the Herbarium of the School of Pharmacy, Second Military Medical University.

*Extraction and Isolation.* The dried stems of *Euonymus alatus* (15 kg) were chopped and extracted at r.t. with 95% EtOH (5×). The extract was concentrated, then diluted with H<sub>2</sub>O and partitioned successively with petroleum ether and AcOEt. The petroleum ether extract (479 g) was subjected to CC (SiO<sub>2</sub>, petroleum ether/acetone 50:1 → 1:1): *Fractions 1–16*. *Fr. 5* (3.4 g) was subjected to CC (*ODS*, MeOH/H<sub>2</sub>O 1:1 → 4:1; then *Sephadex LH-20*, MeOH) and prep. TLC (MeCl/MeOH 30:1): **2** (28.5 mg), **6** (50.2 mg), and **7** (26.1 mg). *Fr. 7* (7.2 g) was purified by CC (*ODS*, MeOH/H<sub>2</sub>O 1:2 → 5:1; then *Sephadex LH-20*, MeOH) and prep. TLC (MeCl/AcOEt/MeOH 8:1:0.1): **1** (11.0 mg), **3** (26.0 mg), **4** (32.2 mg), and **5** (10.9 mg).

rel-[(1*R*,4*S*,4*aS*,7*S*)-1,2,3,4,4*a*,5,6,7-Octahydro-4-hydroxy-7-(1-hydroxy-1-methylethyl)-4*a*-methyl-naphthalen-1-yl]methyl 3-Methylbutanoate (**1**): Colorless oil. [α]<sub>D</sub><sup>20</sup> = +38.0 (*c* = 0.25, MeOH). UV (MeOH): 224. IR (KBr): 3436, 2960, 2871, 1733, 1617, 1467, 1382, 1295, 1191, 1014. <sup>1</sup>H- and <sup>13</sup>C-NMR (CD<sub>3</sub>OD): *Table 1*. ESI-MS: 361 ([*M* + Na]<sup>+</sup>). HR-ESI-MS: 361.2345 ([*M* + Na]<sup>+</sup>, C<sub>20</sub>H<sub>34</sub>NaO<sub>4</sub><sup>+</sup>; calc. 361.2355).

rel-(3*R*,5*S*,5*aR*,6*R*,7*S*,9*S*,9*aS*,10*R*)-10-(Acetyloxy)-5*a*-[(acetyloxy)methyl]octahydro-9-hydroxy-2,2,9-trimethyl-6-(2-methyl-1-oxobutoxy)-7-(1-oxopropoxy)-2*H*-3,9*a*-methano-1-benzoxepin-5-yl Furan-3-carboxylate (**2**): White powder. [α]<sub>D</sub><sup>20</sup> = +40.0 (*c* = 0.25, MeOH). UV (MeOH): 217. IR (KBr): 3445, 2975, 2936, 2879, 1749, 1718, 1462, 1386, 1312, 1245, 1158. <sup>1</sup>H- and <sup>13</sup>C-NMR (CDCl<sub>3</sub>): *Table 2*. ESI-MS: 659 ([*M* + Na]<sup>+</sup>). HR-ESI-MS: 659.2706 ([*M* + Na]<sup>+</sup>, C<sub>32</sub>H<sub>44</sub>NaO<sub>13</sub><sup>+</sup>; calc. 659.2674).

*Evonine* (**3**): Colorless solid. C<sub>36</sub>H<sub>43</sub>NO<sub>17</sub>. <sup>1</sup>H- and <sup>13</sup>C-NMR: matching data in [6]. ESI-MS: 784 ([*M* + Na]<sup>+</sup>). An X-ray diffraction analysis of **3** (*Fig. 5*) established its absolute configuration.

*X-Ray Diffraction Analysis of Evonine (3)*<sup>2</sup>. A single crystal for analysis was obtained from CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O. Data collection was performed with a *Bruker-APEX2-CCD* instrument and graphite-monochromated CuK<sub>α</sub> radiation (λ 1.54178 Å) at 133 (2) K. Crystallographic data: 2 C<sub>36</sub>H<sub>43</sub>NO<sub>17</sub> · 2 MeOH · 3 H<sub>2</sub>O; *M<sub>r</sub>* 1641.56; crystal size 0.20 × 0.18 × 0.16 mm; orthorhombic, space group *P2<sub>1</sub>2<sub>1</sub>2<sub>1</sub>*; *a* = 13.0561 (3) Å, *b* = 13.3515 (3) Å, *c* = 46.4680 (9) Å, α = 90°, β = 90°, γ = 90°; *V* = 8100.2 (3) Å<sup>3</sup>; *Z* = 4; *D<sub>x</sub>* = 1.346 g cm<sup>-3</sup>; *F*(000) = 3480; μ(CuK<sub>α</sub>) 0.934 mm<sup>-1</sup>. Cell refinement and data reduction: The *Bruker SAINT* program was used to solve and refine the structure. *SHELXS-97* and *SHELXL-97*, were used for the refinement on *F*<sup>2</sup> with full-matrix least-squares calculations. All non-H-atoms were filtered with anisotropic parameters, and all H-atoms were positioned by geometrical calculation and refined by the ride-on method with relative isotropic parameters. Absorption correction were applied with semiempirical test from equivalents (max; min. transmission 0.8649;0.8352). In the range θ = 1.90° to

<sup>2</sup>) CCDC-838407 contains the supplementary crystallographic data for this article. These data can be obtained free of charge via [http://www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif).

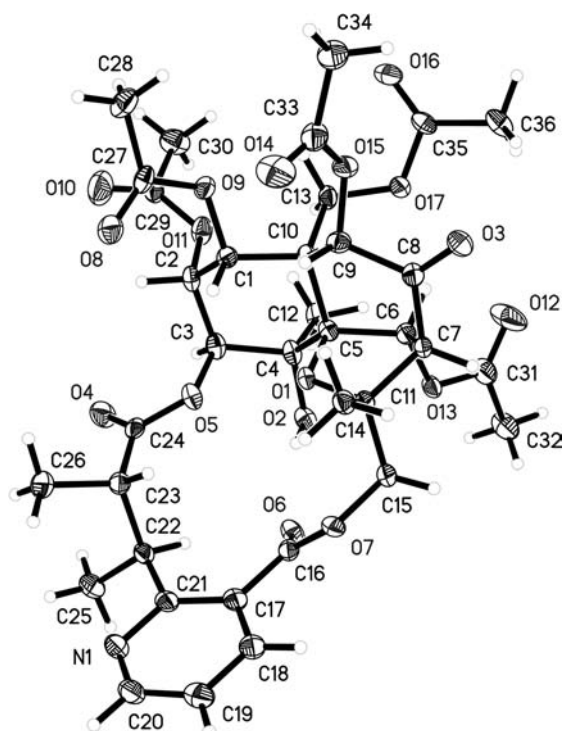


Fig. 5. X-Ray crystal structure of **3** (arbitrary atom numbering)

65.00° and limiting indices  $14 \geq h \geq -13$ , and  $15 \geq k \geq -15$ , and  $54 \geq l \geq -54$ , 57689 reflections (13507 unique,  $R_{\text{int}} = 0.0327$ ) were measured. The final phase converged to  $R_1 = 0.0580$  ( $wR_2 = 0.1617$ ) for 13507 observed reflections ( $I > 2\sigma(I)$ ) and 1040 refined parameters,  $R_1 = 0.0591$  ( $wR_2 = 0.1626$ ) for all unique reflections, and goodness-of-fit on  $F^2$  1.084.

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